



Journal home page: <http://www.journalijar.com>

INTERNATIONAL JOURNAL
OF INNOVATIVE AND APPLIED RESEARCH

RESEARCH ARTICLE

Article DOI: 10.58538/IJAR/2077

DOI URL: <http://dx.doi.org/10.58538/IJAR/2077>

FORMULATION AND CHARACTERIZATION OF POLYHERBAL ANTI-INFLAMMATORY GEL

Shivangi Gupta¹ and Dr. Megha²

1. P.G Scholar of Sirt Pharmacy, Sage University, Bhopal (Madhya Pradesh).
2. Associate Professor, Faculty of Sirt Pharmacy, Sage University, Bhopal (Madhya Pradesh).

Manuscript Info

Manuscript History

Received: 02 January 2024

Final Accepted: 16 February 2024

Published: February 2024

Keywords:

Symplocos Racemosa, Piper Nigrum, Menthol, Capsaicin, Curcuma Amada, and In-Vitro Anti-Inflammatory Properties

Abstract

According to the International Association for the Study of Pain, inflammation is the tissue's immunologic reaction to injury and is characterised by swelling, fluid buildup, and the mobilisation of white blood cells and antibodies. Natural treatments made from plants are called herbal medicines. Instead of eliciting a single molecule that interacts with a single target, they cause a coordinated pharmacological intervention of numerous compounds that interact with multiple targets. Using extracts of Curcuma amada, Piper nigrum, and Lodhra (Symplocos racemosa), menthol, and capsaicin, ten batches of polyherbal gel formulations were created. Formulations F1 through F5 were made with Carbopol 934, and Formulations F6 through F10 were made with HPMC K4M as the gelling agent. The developed formulations were assessed using a range of gel evaluation criteria, including pH determination and physical examination. Measurement of viscosity, washability, extrudability, spreadability, and in vitro diffusion studies. Spreadability of Formulation F2 was good, at 32.48 ± 0.65 g/cm/sec. It was discovered that spreadability decreased when gelling agent concentrations (Carbopol 934 and HPMC K4M) increased. It was discovered that the extrudability of every polyherbal gel formulation ranged from 73.16% to 95.37%. The percentage of medication diffusion decreased as gelling agent concentration increased, according to the results. When compared to polyherbal formulations made with HPMC K4M, it was discovered that the components of formulations made with carbopol 934 were released more readily. The polyphenol content of the gel formulation is responsible for its in vitro anti-inflammatory effect. The stability of the F2 formulation was discovered. The developed formulation must be expanded on a pilot scale, and clinical studies are required to implement the successful introduction of a topical gel formulation including polyherbals for pain relief.

*Corresponding Author: - Shivangi Gupta

Introduction: -

The traditional medical system makes use of several herbs belonging to the Zingiberaceae family. One plant in this family that has been used historically as a stomachic and carminative is curcuma amada, sometimes known as white turmeric [1]. An examination of the literature reveals that these rhizomes contain a variety of chemical

components. Nevertheless, there aren't many sources that assess the extract's pharmacological activity, which includes its carminative, stomachic, and central nervous system effects [2]. The extract demonstrated a substantial inhibitory impact on *Aspergillus niger* and *Trichophyton rubrum*, as well as a hypercholesteremic effect in rabbits. Empirical evidence supports the use of rhizomes as a domestic medicine for inflammatory diseases [3, 4]. *Piper nigrum*, or black pepper, is a member of the Piperaceae family. Native to India, the plants are grown in hot, humid climates [5]. In addition to being used as a spice, black pepper has medicinal properties and is sometimes combined with other well-known herbs and spices to create herbal treatments [6]. The primary physiologically active ingredient in *piper nigrum* is the pungent alkaloid piperine [7]. *Symplocos racemosa* (Symlocaceae), sometimes referred to as "Lodhra" or "Rodhra" in Sanskrit, is a tiny, evergreen tree that can grow up to six metres in height. It can be found in North and East India's plains and lower hills. [8]. The rough, dark grey bark is beneficial for liver problems, diabetes, fever, ulcers, diarrhoea, dysentery, and eye conditions [9]. According to research reports, it possesses antibacterial and anticancer properties and can be effective in treating gynaecological problems [10]. Menthol, sometimes known as "mint camphor," is a popularly accessible volatile oil extract that comes from the plant *Mentha*, which grows mint. Since ancient times, menthol has been applied topically to ease pain [11]. Chili peppers contain a substance called capsaicin, which gives them their fiery and irritating properties. Capsaicin causes pain in addition to the feeling of heat, which makes it a crucial tool for researching pain. One of the main ingredients in spicy peppers, capsaicin, was thought to have anti-inflammatory properties [12].

In the global health services system, these traditional medicines play a vital role. Opioids, also known as non-steroidal anti-inflammatory medicines, are commonly used to treat a variety of types of inflammation. However, they have serious adverse effects, including itching and redness. Therefore, it appears that looking for more advantageous options would be necessary. Gel formulations are preferred over other topical preparations and oral administration for drug delivery topically because they are simple to apply, extend contact time, and cause fewer side effects [13]. The goal of the current work was to create a polyherbal gel with anti-inflammatory properties *in vitro*.

Material And Methods:-

Materials:-

We bought dried *Curcuma amada* rhizomes, dried *Piper nigrum* fruits, and dried *Lodhra* (*Symplocos racemosa*) bark from the Nashik local market. Prof. Manohar Gulab Gavit of the Department of Botany at the MVPS KANMS Arts, Commerce, and Science College in Nashik, Maharashtra, verified the authenticity of the plant materials. (Herbarium 3; Authentication No. KANMS/2020-21/56). Naturite Agro Products Limited, Hyderabad, India, sent a free sample of capsaicin. We bought menthol from S. D. Fine Chemicals in Mumbai. We purchased HPMC K4M and carbopol 934 from N.R. Chem in Bombay. Analytical-grade compounds were employed for all other substances.

Methods:-

The process of making plant extract

In a prior study, the authors conducted and assessed methanolic extracts of black pepper (*Piper nigrum*) fruits (MEPN), white turmeric (*Curcuma amada*) rhizome (EECA), and *lodhra* (*Symplocos racemosa*) bark (MESR) for different phytochemical screenings. These extracts were used along with carbopol 934 and HPMC K4M as gelling agents to generate a polyherbal gel.

Formulation of Polyherbal Gel:

Preparation of gel base Schmolka et al.'s cold mechanical technique was used to create the gel base. To prevent agglomeration, carbopol 934/HPMC K4M was dissolved gradually and stirred in 60 mL of demineralized water for one hour. After that, 10 mL of demineralized water was used to dissolve triethanolamine and disodium edetate, and the mixture was agitated for ten minutes. For ten minutes, 4.83 ml of propylene glycol was stirred with 12 mL of demineralized water. Carbopol 934/HPMC K4M solution was mixed with triethanolamine solution and sodium edetate, and the mixture was stirred for ten minutes to bring the pH down to 7.4. Subsequently, a clear, uniform gel basis was created by adding propylene glycol solution and stirring for ten minutes [14].

Preparation of gel formulation

Ten formulations with varying extract concentrations (EECA, MEPN, and MESR) were made. Formulations F1 through F5 were made with Carbopol 934, and Formulations F6 through F10 were made with HPMC K4M as the gelling agent. (Table 1).

Drugs such as EECA, MEPN, MESR, menthol, and capsaicin were added to the gel base and stirred continuously until the drugs were fully dissolved. The aluminium collapsible tube was filled with the prepared gel and sealed. For the base control gel without the extract and additional active component, a similar process was used [14, 15, 16].

Evaluation of Polyherbal Topical Gel Physical Examination

After being packaged, each gel was visually inspected to ensure homogeneity. Their appearance and the existence of any aggregates were examined [17].

Determination of pH

Using a pH metre, the pH of each gel composition was determined. Before being used, the pH metre was calibrated using standard buffer solutions at pH 4, 7, and 9. The electrode was placed in the sample at room temperature for 10 minutes prior to obtaining the reading, and the pH was then recorded [18, 19, 20].

Viscosity Measurement

Using Brookfield's rheometer (R/S-CPS+ rheometer with C75-2 measurement system) at 30 rpm and room temperature ($25\pm 2^\circ\text{C}$), the viscosity of the gel formulations (0.5 g each) was measured [21].

Spreadability

The device, which was altered, is made up of a wooden block with a pulley at one end. On the block, a rectangular ground glass plate was mounted. Gel (around 2 grammes) was put on the bottom plate and sandwiched between the hook-equipped lower and upper glass plates, which were of the same size. To get a homogenous gel film and to release any trapped air, a 500 mg weight was placed on top of each of the two plates for five minutes. Extra gel was removed with a scraper. A 50-gram pull was applied to the upper plate. The upper plate's needed time (in seconds) to travel

10 centimetres was recorded. The spreadability was determined using the subsequent formula. Spreadability is better when the time gap is shorter [22].

$S = M \times L/T$, where M is the weight attached to the upper slide, L is the glass slide's length, and T is the amount of time it takes for plates to travel the whole length (in seconds).

Extrudability

Standard-capped collapsible aluminium tubes were filled with the gel compositions, and the ends were crimped shut to seal. It was noted how much each tube weighed. The tubes were clamped after being positioned between two glass slides. After covering the slides with 0.5 g, the cap was taken off. Weighing was done on the amount of extruded gel that was collected. It was determined what the ~~what~~ percentage of the extruded gel was [23, 24].

Grittiness

Every formulation was examined under a light microscope to determine whether any significant particle matter was present. Therefore, it is evident that the gel preparation satisfies the necessary conditions of being devoid of specific material and forming the requisite grittiness for any topical preparation [20, 24, 25, 26].

Washability

After applying formulations to the skin, the degree and simplicity of water washing were personally assessed [27].

In-vitro diffusion study

Using a traditional standard cylindrical tube that was made in the lab using a straightforward modification of a glass tube with an internal diameter of 15 mm and a height of 100 mm, the in-vitro diffusion of all polyherbal formulations and commercialised Diclofenac sodium gel 0.5% was investigated. A donor compartment was created by tying the commercial semi-permeable membrane cellophane, which had been presoaked overnight in the freshly made dissolving medium (phosphate buffer pH 6.8), to one end of an open cylinder that was open on both sides. One gramme of the polyherbal formulation was inserted into this slot. The membrane of the diffusion cell served as the corneal epithelium. All of the membrane's surface came into contact with the receptor compartment, which held 25 millilitres of phosphate buffer (pH 6.8) in a 100-milliliter beaker. A magnetic stirrer was used to continually swirl the contents of the receptor compartment, and a temperature of 37°

$\pm 0.5^{\circ}\text{C}$ was maintained. One millilitre of the sample (aliquot) was taken out of the receptor compartment and replaced with a new buffer solution at predetermined intervals. After making the necessary dilutions against reference using phosphate buffer pH 6.8 as a blank, the aliquot was examined for drug concentration using a UV-VIS spectrophotometer at a wavelength that was determined [14, 28].

Selection of Satisfactory Formulation of Polyherbal Gel

The selection of 10 polyherbal gel formulations was based on many factors, such as pH, viscosity, homogeneity, extrudability, washability, and homogeneity.

Evaluation of Satisfactory formulation Thixotropy Analysis

Using Brookfield's rheometer (R/S-CPS+ rheometer with C75-2 measuring system) at 30 rpm and room temperature ($25\pm 2^{\circ}\text{C}$), the viscosity of ordinary Lisinopril gel (0.5g) and Lisinopril ion-pair gel of the F2 formulation was evaluated [29].

Table 2:- Evaluation Of Polyherbal Gel.

<u>Formulation</u>	<u>Consistency</u>	<u>Ph</u>	<u>Viscosity</u> <u>(Pa.S)</u>	<u>Spreadibility</u> <u>(g.cm/sec)</u>	<u>Homogeneity</u>	<u>Washability</u>
F1	Fluid	<u>6.22±0.13</u>	<u>4.86±0.89</u>	<u>45.58±1.56</u>	Homogeneous	Excellent
F2	Semisolid	<u>6.84±0.38</u>	<u>7.61±0.34</u>	<u>32.48±0.65</u>	Homogeneous	Excellent
F3	Semisolid	<u>6.35±0.43</u>	<u>8.97±0.66</u>	<u>24.73±1.08</u>	Homogeneous	Good
F4	Semisolid	<u>6.43±0.19</u>	<u>10.42±0.91</u>	<u>18.94±1.44</u>	Homogeneous	Good
F5	Stiff	<u>6.73±0.44</u>	<u>12.63±1.06</u>	<u>12.38±1.93</u>	Homogeneous	Good
F6	Fluid	<u>6.38±0.15</u>	<u>3.96±0.26</u>	<u>46.31±1.42</u>	Homogeneous	Excellent
F7	Fluid	<u>6.86±0.42</u>	<u>4.53±0.43</u>	<u>38.49±1.81</u>	Homogeneous	Excellent
F8	Semisolid	<u>6.72±0.39</u>	<u>6.73±1.28</u>	<u>31.92±1.33</u>	Homogeneous	Good
F9	Semisolid	<u>6.64±0.23</u>	<u>9.68±1.08</u>	<u>25.66±1.27</u>	Homogeneous	Good
F10	Stiff	<u>6.82±0.67</u>	<u>11.39±1.44</u>	<u>21.34±1.08</u>	Homogeneous	Good

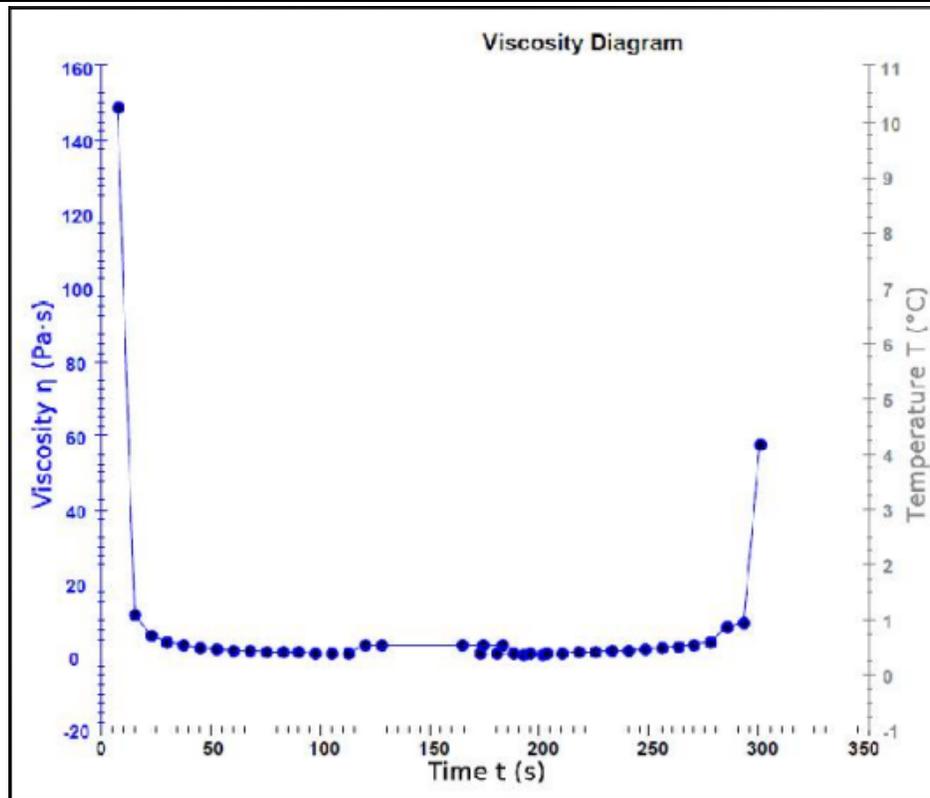


Fig. 2:- Viscosity diagram.

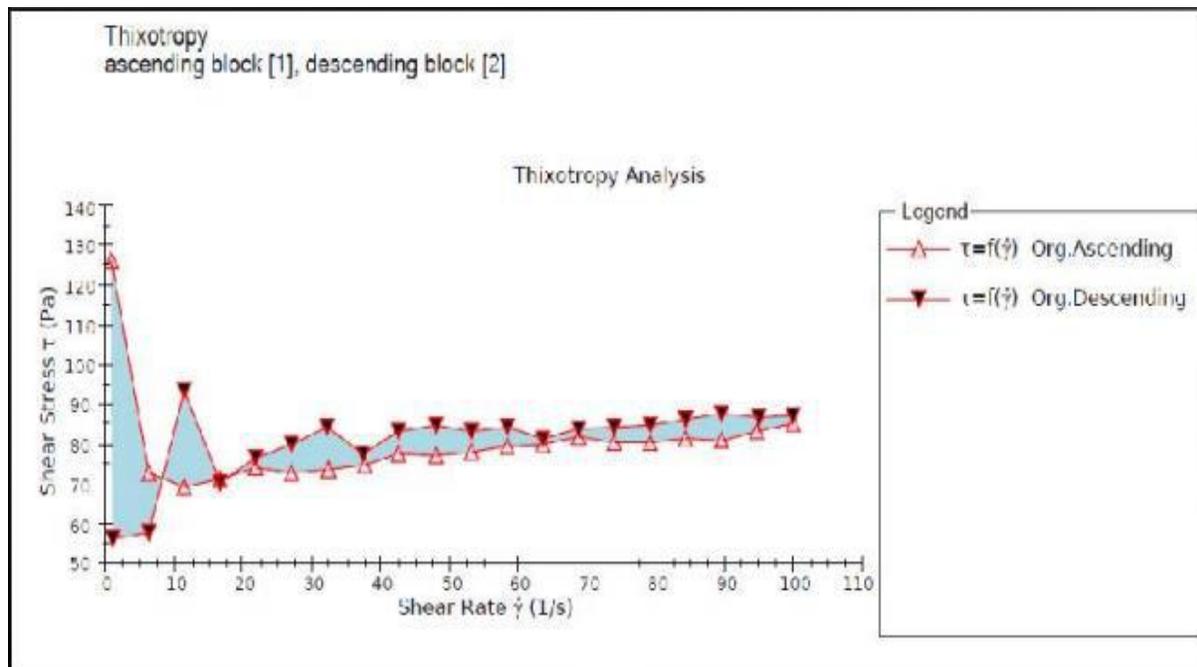


Fig. 3:- Thixotropy analysis.

Stability Studie

Stability studies were carried out in accordance with ICH recommendations on F2 formulation since it showed superior quality features in order to guarantee the quality of polyherbal gel formulation throughout its shelf life. Viscosity, pH, and homogeneity all showed negligible changes during 0, 1, 2, 3, and 6 months of stability testing. The study's findings unequivocally demonstrated the stability of the topical polyherbal gel F2.

In-vitro Anti-Inflammatory Inhibition of albumin denaturation

To explore the mechanism behind the anti-inflammatory action, the extract's capacity to prevent protein denaturation was assessed. This approach to assessing anti-inflammatory activity is easy to use and straightforward. The results of this study showed that the polyherbal gel formulation F2 and the reference drug Diclofenac sodium showed concentration-dependent suppression of protein denaturation in the range of 10–50 $\mu\text{g/ml}$. A maximum inhibition of

82.40% was noted at 50 $\mu\text{g/ml}$ of the F2 formulation. At a concentration of 50 $\mu\text{g/ml}$, diclofenac sodium exhibited the highest level of inhibition, measuring 85.47%.

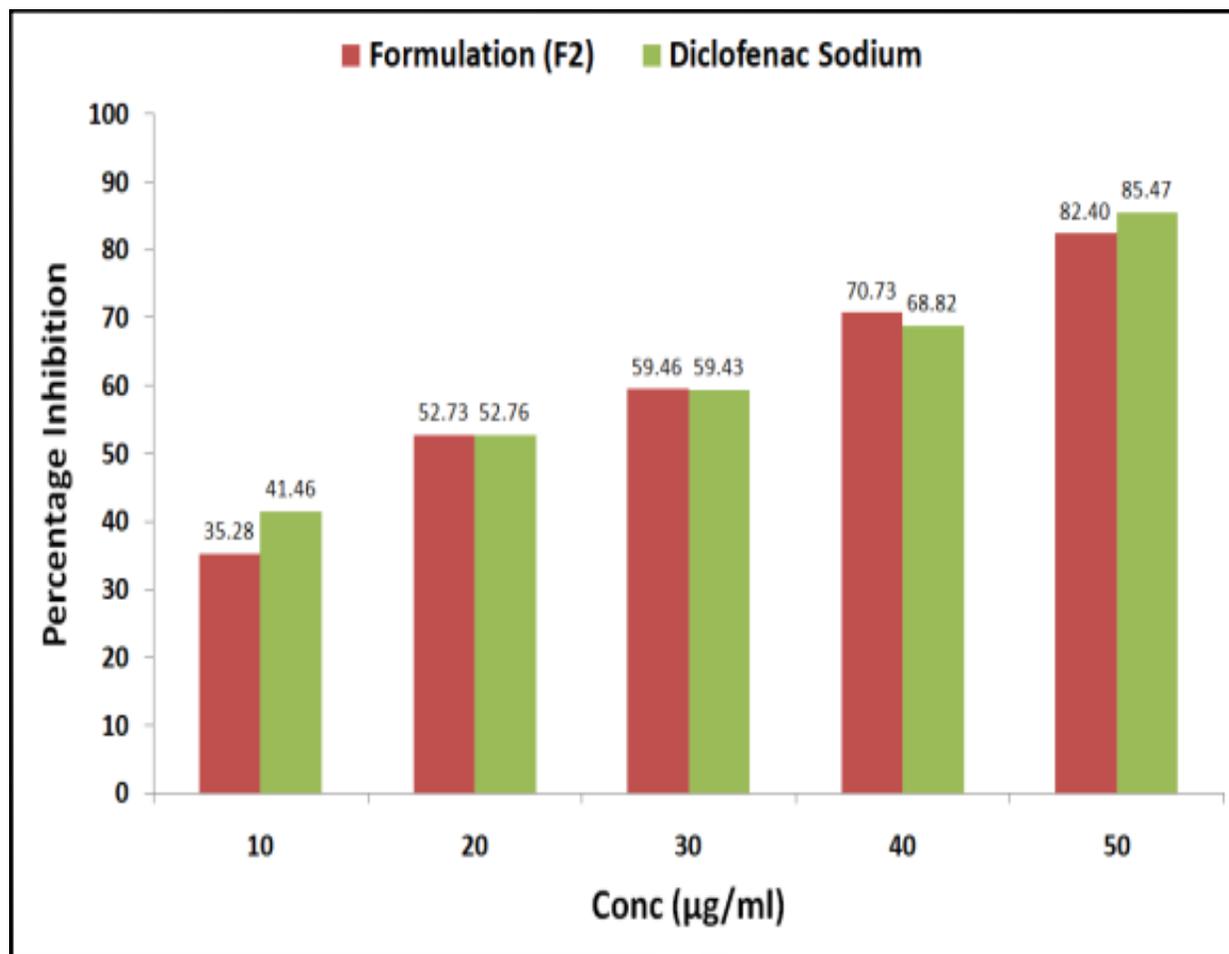


Fig. 4:- Effect of polyherbal formulation (F2) on heat-induced protein denaturation.

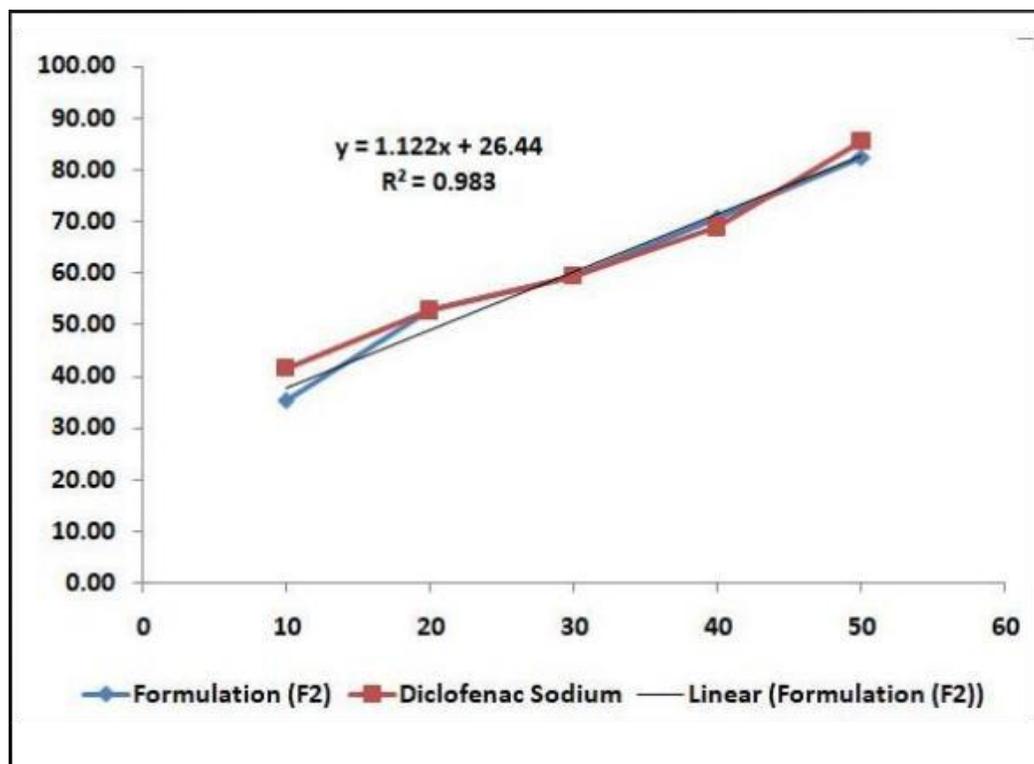


Fig. 5:- Correlation between % inhibition of formulation (F2) and Diclofenac sodium.

Conflict Of Interest

The writers affirm that there is no conflict of interest between them.

References:-

- Hussain A, Virmani OP, Popli SP, Misra, LN, Gupta MM et al. (1992). Dictionary of Indian medicinal plants. Director, Central Institute of Medicinal and Aromatic Plants, Lucknow, 161-2.
- Jain MK, Mishra RK. (1964). Chemical examination of Roxb. Indian J Chem. 2:39.
- Gholap AS, Bandyopadhyay C. (1984). Characterization of mangolike aroma in *C. amada* Roxb. J Agric Food Chem.; 32:7-9.
- Mujumdar A.M., Naik D.G., Dandge C.N., and Puntambekar H.M. (2000). Antiinflammatory Activity of Curcuma Amada Roxb. in Albino Rats. Indian Journal of Pharmacology, 32: 375-377.
- Hamrapurkar PD, Jadhav Kavita, and Zine Sandip. (2011). Quantitative estimation of piperine in *Piper nigrum* and *Piper longum* using high-performance thin-layer chromatography. Journal of Applied Pharmaceutical Sciences, 01(03): 117-120.
- Sharma Pradeep Kumar, et al. (2013). Evaluation of *Zingiber officinale* and *Curcuma longa* rhizome as crude drugs from their ethanolic extract. Int. Res. J. Pharm. 4(12): 2230-8407.
- Wood AB, Maureen L., Barrow, and James DJ. (1988). Piperine determination in pepper (*Piper nigrum* L.) And its oleoresins: A reversed-phase high-performance liquid chromatographic method, flavour and fragrance Journal, 3: 55-64.
- Sharma, PC. (2002). Data Base on Medicinal Plants Used in Ayurveda. Central Council for
- Research in Ayurveda and Siddha, 5:164-168.
- Kumar GS, Jayaveera KN, Ashok Kumar CK, Umachigi PS, Vrushabendra BM, and Kishore DV. (2007). Antimicrobial effects of Indian medicinal plants against acne-inducing bacteria. Tropical J Pharma Res. 6:717-23.
- Sharma SK, Sharma SM, Saini V, and Mohapatra S. (2013). Evaluation of Analgesic and Anti-Inflammatory Activity of *Symplocos Racemosa*. International Research Journal of Pharmacy, 4(2): 136-139.

13. Pergolizzi JV., et al. (2018). The role and mechanism of action of menthol in topical analgesic products. *Clin Pharm Ther.* 43:313–319.
14. Victor Fattori, et al. (2016). Capsaicin: Current Understanding of Its Mechanisms and Therapy of Pain and Other Pre-Clinical and Clinical Uses. *Molecules*, 21:844, 1-33.
15. Dixit G., Misal G., Gulkari V., and Upadhye K. (2013). Formulation and Evaluation of Polyherbal Gel for Anti-Inflammatory Activity. *Int J Pharm Sci Res.* 4(2); 1186–1191.
16. Rajasekaran Aiyalu, Arulkumaran Govindarjan, and Arivukkarasu Ramasamy. (2016).
17. Formulation and evaluation of topical herbal gel for the treatment of arthritis in an animal model. *Brazilian Journal of Pharmaceutical Sciences*, 52(3): 493-507.
18. Schmolka, IR. (1972). Preparation and properties of Pluronic PF-127 gels for the treatment of burns. *J Biomed Mater Res.*; 6:571–82.
19. Divya Jyothi, Marina Koland. (2015). Formulation and Evaluation of an Herbal
20. Anti-Inflammatory Gel Containing *Trigonella foenum greacum* seed extract. *Int J Pharm Pharm Sci.* 8(1): 41–44.
21. Rajan R., Vasudevan D. (2012). Effect of permeation enhancers on the penetration mechanism of transdermal gel of ketoconazole. *J. Adv. Pharma. Tech. Res.*;3(7):112-116.
22. Queiroz, M.B.R.; Marcelino, N.B.; Ribeiro, M.V.; Espindola, L.S.; Cunha, F.; Silva, M.V. (2009).
23. Development of gel with *Matricaria recutita* L. extract for topical application and evaluation of physical-chemical stability and toxicity. *Lat. Am. J. Pharm.*; 28(4): 574-579.
24. Basha BN, Prakasam K, and Goli D. (2011). Formulation and evaluation of gel containing the fluconazole-antifungal agent. *Int J Drug Dev Res.* 3:109–28.
25. Jyothi D., Koland M. (2015). Formulation and Evaluation of an Herbal Anti-Inflammatory Gel Containing *Trigonella foenum greacum* seed extract. *Int J Pharm Pharm Sci.* 8(1): 41–44.
26. Mulani H., Bhise K. (2017). QbD Approach the formulation and evaluation of miconazole-nitrate-loaded ethosomal cream-o-gel. *Int. Res. J. Pharm. Sci.* 8:1-37.
27. Cui H., Quan P., Zhou Z., and Fang L. (2016). Development of a drug-in-adhesive patch combining ion pairs and a chemical enhancer strategy for topical delivery of zaltoprofen: pharmacokinetics and in vitro/in vivo correlation evaluation. *Drug Deliv.* 23(9):3461-3470.
28. Sudipta D., Haldar PK, and Pramanik G. (2011). Formulation and evaluation of herbal gel containing *Clerodendrum infortunatum* leaf extract. *Int J Pharmtech Res.* 3:140–3.
29. Giri MA, Bhalke RD (2019). Formulation and Evaluation of Topical Anti-Inflammatory Herbal Gel. *Asian J Pharm Clin Res.* 12(7): 252-255.
30. Goyal S, Sharma P, Ramchandani V, Shrivastava SK, and Dubey PK. (2011). Novel anti-inflammatory topical herbal gels containing *Withania somnifera* and *Boswellia serrata*. *Int J Pharm Biol Sci Arch.* 2:1087-94.
31. Sharma M., Rathore V. (2014). Formulation, Development, and Evaluation of Novel
32. Poly-Herbal Anti-Acne Gel. *Int.J.PharmTech Res.* 6(1): 58–62.
33. Susheel Thakur, Nisha Thakur, and Niladry Shekar Ghosh. (2016). Formulation and in-vitro evaluation of a polyherbal micro-emulgel containing *Tinospora cordifolia* and curcumin for the treatment of arthritis. *International Journal of Pharmaceutical Sciences and Drug Research*, 8(5): 259–264.
34. Mishra US, Murthey PN, Mishra D, and Sahu K. (2011). Formulation and stabilization of herbal gel containing methanolic extract of *Calophyllum inophyllum*. *Am J Pharmtech Res.* 1:276–89.
35. Priscilla R. Varges, Camila M. Costa, Bruno S. Fonseca, Mônica F. Naccache, and Paulo R. de Souza Mendes. (2019). Rheological Characterization of Carbopol® Dispersions in Water and in Water/Glycerol Solutions. *Fluids*, 4, 3.
36. Singh M., Mittal V. (2014). Formulation and evaluation of a herbal gel containing an ethanolic extract of *Ipomoea fistulosa*. *Int J Sci Res.* 3:25–9.
37. Sakat S, Juvekar AR, and Gambhire MN. (2010). In vitro antioxidant and anti-inflammatory activity of methanol extract of *Oxalis corniculata* Linn. *Int J Pharma and Pharmacol Sci.* 1.
38. Gautam RK, Sharma S, and Sharma K (2013). Comparative evaluation of anti-arthritic activity of *Pongamia pinnata* (Linn.) Pierre and *Punica granatum* An in-vitro study. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(4), 721–724.
39. Shallangwa GA, Musa H, and Nyaga GT (2015). In-vitro evaluation of ethanolic extracts of *Zingiber officinale* *Syzygium aromaticum* and their 1:1 extract blend on protein denaturation during acute inflammation. *Journal of Progressive Research in Chemistry*, 1(1):1–8.
40. Imam S, Shaheen N, Tasleem F, Azhar I, and Mahmood ZA (2017). Evaluation of anti-inflammatory and other biological activities of flavonoid-based cream formulations for topical application using in vitro model. *Int J Pharm Sci Res.* 8(10): 4388–95.