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## RESEARCH ARTICLE

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## Neutrophil Dynamics and Host Defense Mechanisms in Pregnant Women Infected with Trichomonas vaginalis

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#### Abstract

..... Trichomonas vaginalis, a prevalent sexually transmitted parasite, poses substantial risks to maternal health during pregnancy, eliciting a multifaceted immune response crucial for host defense. Neutrophils, as primary innate immune effectors, play a pivotal role in combatting this infection within the intricate immunological landscape of pregnancy. This comprehensive review aims to elucidate the complex interplay between Trichomonas vaginalis infection and the dynamic responses of neutrophils in pregnant women, exploring mechanisms of neutrophil recruitment, activation, effector functions, and the parasite's evasion strategies. Insights into neutrophil dynamics and activation mechanisms reveal their essential functions in combating T. vaginalis, encompassing chemotaxis, phagocytosis, release of reactive oxygen species, and formation of neutrophil extracellular traps. Furthermore, the review discusses how pregnancy-associated immunomodulation influences neutrophil function in response to this parasitic infection. Concurrently, the elucidation of T. vaginalis evasion tactics-surface antigen variation, adhesion strategies, and immune subversion-underscores the complexity of host-parasite interactions and the challenges faced by neutrophils in eradicating the parasite. Considering the clinical implications, particularly adverse pregnancy outcomes and maternal morbidity associated with T. vaginalis infection, the review addresses current treatment modalities, management challenges during pregnancy, and potential therapeutic strategies targeting responses and immune modulation. In conclusion, neutrophil understanding the intricate interplay between T. vaginalis infection and neutrophil dynamics within pregnancy's immunological context provides valuable insights into potential therapeutic targets. This review advocates for further research aiming to enhance our understanding of neutrophilparasite interactions and develop targeted interventions to ameliorate adverse outcomes associated with T. vaginalis infection in pregnant women.

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### **Introduction:-**

Trichomonas vaginalis, a flagellated protozoan parasite, stands as one of the most prevalent sexually transmitted infections globally, exerting significant implications for maternal health during pregnancy [1]. Among the myriad of complications that afflict expectant mothers, the interplay between this parasitic infection and the intricate immunological landscape of pregnancy unveils a captivating yet complex relationship. Neutrophils, as primary innate immune cells, emerge as key orchestrators in combating Trichomonas vaginalis, shaping the host defense mechanisms within this unique physiological state [2-3]. The epidemiology of Trichomonas vaginalis during pregnancy portrays an alarming prevalence, underscoring its potential impact on maternal health and pregnancy outcomes. Heightened susceptibility to this parasite during gestation, attributed to hormonal, anatomical, and immunological changes, demands a thorough exploration of the immune responses elicited, with a particular focus on the role of neutrophils within this specialized immunological milieu [4]. Neutrophils, traditionally regarded as the first line of defense against invading pathogens, manifest a multifaceted response upon encountering Trichomonas vaginalis. Their dynamic interplay includes chemotaxis to the site of infection, phagocytosis, degranulation, and the formation of neutrophil extracellular traps (NETs), aiming to eliminate the parasite and mitigate its pathological consequences. However, the parasite's evasion strategies, including surface antigen variation and immune modulation, present formidable challenges to neutrophil-mediated eradication, emphasizing the complexity of hostpathogen interactions [5-6]. Moreover, the immunomodulatory effects inherent to pregnancy impart significant alterations in neutrophil function and immune responses, potentially influencing the host defense mechanisms against Trichomonas vaginalis. Understanding these modifications within the pregnant immunological milieu is pivotal in delineating the nuances of neutrophil dynamics and their efficacies in combating this parasitic infection.

### **Neutrophil Dynamics and Activation**

Neutrophils, as pivotal innate immune effectors, orchestrate a multifaceted response against Trichomonas vaginalis, the causative agent of one of the most prevalent sexually transmitted infections globally. Within the distinctive immunological milieu of pregnancy, neutrophils undergo dynamic changes in their activation, recruitment, and effector functions in response to Trichomonas vaginalis, contributing significantly to the host defense mechanisms [7-10].Upon encountering Trichomonas vaginalis, a cascade of signaling events triggers the recruitment of neutrophils to the site of infection. Chemotactic signals, including cytokines, chemokines, and damage-associated molecular patterns (DAMPs), guide the migration of neutrophils from the circulation to the infected tissues, aiming to contain and eliminate the parasite [11].Neutrophils employ phagocytosis as a primary mechanism to engulf and internalize Trichomonas vaginalis, initiating intracellular killing processes. This process involves the formation of phagosomes followed by fusion with lysosomes, releasing antimicrobial peptides, proteases, and reactive oxygen species (ROS) to eliminate the parasite. [12] Activation of neutrophils in response to Trichomonas vaginalis triggers the release of Neutrophil Extracellular Traps (NETs)-a web-like structure comprising DNA, histones, and antimicrobial proteins-to ensnare and neutralize the parasite. However, the parasite's mechanisms to evade NETmediated killing pose challenges to the efficacy of this neutrophil effector mechanism [13]. Neutrophils generate Reactive Oxygen Species (ROS), such as superoxide anions and hydrogen peroxide, as part of their antimicrobial arsenal against Trichomonas vaginalis. These reactive molecules exert cytotoxic effects, aiding in the eradication of the parasite. However, the delicate balance between ROS-mediated killing and potential tissue damage underscores the complexity of neutrophil responses [14-16]. The unique immunological milieu of pregnancy exerts modulatory effects on neutrophil function and responses. Hormonal fluctuations, altered cytokine profiles, and changes in immune cell populations during gestation influence neutrophil activation and their ability to mount an effective response against Trichomonas vaginalis [17-18]. The intricate dynamics of neutrophil activation and effector functions in response to Trichomonas vaginalis underscore the complexities of host-pathogen interactions within the pregnant immunological landscape. Understanding these mechanisms is critical in devising targeted interventions to enhance neutrophil-mediated immunity and mitigate adverse outcomes associated with T. vaginalis infection during pregnancy.

### **Parasite Evasion Mechanisms and Immune Subversion**

Trichomonas vaginalis, a prevalent sexually transmitted parasite, employs an array of sophisticated evasion strategies to subvert host immune responses, presenting formidable challenges to effective immune clearance. particularly within the distinctive immunological milieu of pregnancy. Understanding the parasite's evasion mechanisms is pivotal in elucidating the complexities of host-parasite interactions and devising strategies to counteract immune subversion [19-20]. Trichomonas vaginalis exhibits remarkable phenotypic plasticity through the expression of variable surface proteins (VSPs), enabling rapid antigenic variation. This antigenic switching mechanism allows the parasite to evade host immune recognition, facilitating persistence and chronic infection, particularly within the vaginal mucosa during pregnancy [21-24]. The parasite's adherence to host cells, mediated by surface adhesins and lectins, not only facilitates colonization but also serves as a mechanism to evade immune detection. Trichomonas vaginalis modulates host immune responses by inducing an immunosuppressive microenvironment, hindering effective neutrophil responses and impairing the activation of other immune effectors [25].Trichomonas vaginalis exerts immunomodulatory effects by influencing cytokine profiles, altering immune cell activation, and dampening inflammatory responses. By disrupting the balance between pro-inflammatory and antiinflammatory mediators, the parasite manipulates immune signaling pathways, thereby evading immune recognition and clearance mechanisms [26]. Trichomonas vaginalis possesses robust antioxidant defense mechanisms, including enzymes such as superoxide dismutase and peroxiredoxins, enabling the parasite to counteract host-derived oxidative stress. This contributes to the parasite's resilience against neutrophil-mediated killing and aids in immune evasion strategies [27]. Trichomonas vaginalis deploys mechanisms to evade neutrophil-mediated killing, including resistance to Neutrophil Extracellular Trap (NET)-induced toxicity or degradation by nucleases released by the parasite. This evasion strategy diminishes the efficacy of neutrophil defense mechanisms and contributes to the parasite's persistence [28]. Trichomonas vaginalis demonstrates adaptability to the specific immunological changes associated with pregnancy. The altered hormonal milieu and immunomodulatory effects during gestation potentially influence the parasite's survival strategies, complicating the host immune responses and perpetuating the infection [29].The elucidation of Trichomonas vaginalis evasion tactics and immune subversion strategies provides critical insights into the challenges faced by the host immune system, particularly during pregnancy. Understanding these mechanisms is crucial in developing targeted therapeutic interventions to counteract parasite evasion, mitigate adverse outcomes, and enhance immune clearance of T. vaginalis in pregnant women.

#### **Clinical Implications and Therapeutic Perspectives**

Trichomonas vaginalis infection during pregnancy poses significant clinical implications, necessitating tailored therapeutic approaches to mitigate adverse outcomes and ensure optimal maternal and fetal health. Understanding the challenges in managing this prevalent parasitic infection within the context of pregnancy is pivotal for developing effective therapeutic strategies and improving clinical outcomes [30-40]. Trichomonas vaginalis infection during pregnancy is associated with various adverse outcomes, including preterm birth, low birth weight, premature rupture of membranes, and increased susceptibility to other sexually transmitted infections. These complications underscore the importance of timely diagnosis and management strategies to minimize risks to maternal and neonatal health [41-52]. The management of Trichomonas vaginalis infection during pregnancy poses unique challenges, primarily due to safety concerns associated with the use of antiparasitic medications in expectant mothers. Limited therapeutic options approved for use during pregnancy necessitate careful consideration of risks and benefits in choosing appropriate treatment regimens [53-62]. Routine screening for Trichomonas vaginalis infection during prenatal care remains crucial for early detection and timely intervention. Incorporating accurate diagnostic methods, such as nucleic acid amplification tests or point-of-care testing, into antenatal care protocols can facilitate prompt identification and treatment of infected individuals [63-74]. Tailoring treatment strategies involves considering safety profiles and efficacy of available medications during pregnancy. Nitroimidazoles, such as metronidazole and tinidazole, are commonly used, but their use in the first trimester requires cautious evaluation due to potential teratogenic effects [75-83]. Exploring combination therapies or alternative treatment approaches, including topical agents or adjunctive therapies, warrants consideration to enhance therapeutic efficacy and minimize potential risks associated with single-agent regimens [84-90]. Implementing preventive measures, such as promoting safe sexual practices and partner treatment, along with patient education, is pivotal in preventing recurrent infections and reducing the risk of transmission [91-96]. Continued research endeavors focusing on the safety and efficacy of antiparasitic agents, the development of novel therapeutics targeting parasite virulence factors, and the assessment of vaccine candidates against Trichomonas vaginalis infection are essential for advancing treatment options for pregnant women. Strategizing comprehensive care approaches that encompass screening, safe and effective treatment options, preventive measures, and patient education are critical in mitigating the clinical implications of Trichomonas vaginalis infection in pregnant women. Advancing research efforts and developing

targeted therapeutic interventions hold promise for improving maternal and neonatal outcomes in the context of this prevalent parasitic infection during pregnancy [97-100].

# **Conclusion:-**

Trichomonas vaginalis infection poses a significant health concern for pregnant women, presenting challenges in diagnosis, treatment, and potential adverse outcomes. Within the unique immunological context of pregnancy, the intricate interplay between the parasite's evasion mechanisms and the host immune responses underscores the complexities of managing this infection during gestation. Neutrophils, as primary innate immune effectors, play a pivotal role in combating Trichomonas vaginalis; however, the parasite's ability to evade immune clearance through surface antigen variation, adhesion strategies, and immune subversion presents formidable challenges to effective clearance mechanisms, especially in pregnant women. The clinical implications of T. vaginalis infection during pregnancy, including adverse birth outcomes and challenges in treatment, emphasize the necessity for tailored therapeutic approaches. The limited safety data regarding antiparasitic medications in pregnancy necessitates a cautious approach in selecting treatment regimens, while continual research efforts are crucial to expand therapeutic options and enhance efficacy while minimizing risks. Early screening, accurate diagnosis, preventive measures, and patient education play critical roles in mitigating the impact of Trichomonas vaginalis infection in pregnant women. Advances in diagnostic techniques, exploration of alternative treatment modalities, and the development of preventive strategies are pivotal for improving clinical outcomes and reducing the burden of this infection on maternal and neonatal health. A multifaceted approach encompassing early detection, safe and effective treatment options, preventive measures, and ongoing research initiatives is essential in addressing the clinical implications of Trichomonas vaginalis infection during pregnancy. Enhancing our understanding of host-parasite interactions and advancing therapeutic strategies tailored to the unique immunological environment of pregnancy holds promise for improving maternal and neonatal outcomes in the context of this prevalent parasitic infection.

## **References:-**

- 1. Van Der Pol B. Trichomonas vaginalis. Diagnostics to Pathogenomics of Sexually Transmitted Infections. 2018:342.
- 2. Hamouda MM, Mohamed SA, Nabih N, El-Henawy AA, Eldeen NE, EL-zayady WM. Trichomonas vaginalis infection and pregnancy outcome.
- 3. Ochei KC, Obeagu EI, Ugwu GU, George CN (2014). Prevalence Of Trichomonas Vaginalis Among Pregnant Women Attending Hospital in Irrua Specialist Teaching Hospital in Edo State, Nigeria. IOSR Journal of Dental and Medical Sciences (IOSR-JDMS), 2014; 13(9 Ver. II):79-82
- 4. Galani IE, Andreakos E. Neutrophils in viral infections: current concepts and caveats. Journal of Leucocyte Biology. 2015;98(4):557-64.
- 5. Kobayashi SD, Malachowa N, DeLeo FR. Neutrophils and bacterial immune evasion. Journal of Innate Immunity. 2018;10(5-6):432-41.
- 6. Kumar M, Saadaoui M, Al Khodor S. Infections and pregnancy: Effects on maternal and child health. Frontiers in Cellular and Infection Microbiology. 2022:690.
- 7. Mielczarek E, Blaszkowska J. Trichomonas vaginalis: pathogenicity and potential role in human reproductive failure. Infection. 2016; 44:447-58.
- 8. Dunne RL, Dunn LA, Upcroft P, O'donoghue PJ, Upcroft JA. Drug resistance in the sexually transmitted protozoan Trichomonas vaginalis. Cell research. 2003;13(4):239-49.
- V'kovski P, Kratzel A, Steiner S, Stalder H, Thiel V. Coronavirus biology and replication: implications for SARS-CoV-2. Nature Reviews Microbiology. 2021;19(3):155-70.
- Amala SE, Karibi-Botoye R, Nwokah EG, Pius MT. Prevalence of Asymptomatic Bacteriuria in Pregnancy and Urinary Tract Infection in Non-pregnant Symptomatic Womem. American Journal of Biomedical Sciences. 2021;13(4).
- 11. Silver BJ, Guy RJ, Kaldor JM, Jamil MS, Rumbold AR. Trichomonas vaginalis as a Cause of Perinatal Morbidity. Sexually transmitted diseases. 2014;41(6):369-76.
- 12. Kigozi GG, Brahmbhatt H, Wabwire-Mangen F, Wawer MJ, Serwadda D, Sewankambo N, Gray RH. Treatment of Trichomonas in pregnancy and adverse outcomes of pregnancy: a subanalysis of a randomized trial in Rakai, Uganda. American journal of obstetrics and gynecology. 2003;189(5):1398-400.
- 13. Mabey D, Ackers J, Adu-Sarkodie Y. Trichomonas vaginalis infection. Sexually transmitted infections. 2006;82(suppl 4): iv26-7.

- 14. Okun N, Gronau KA, Hannah ME. Antibiotics for bacterial vaginosis or Trichomonas vaginalis in pregnancy: a systematic review. Obstetrics & Gynecology. 2005;105(4):857-68.
- 15. Marrazzo JM, Coffey P, Bingham A. Sexual practices, risk perception and knowledge of sexually transmitted disease risk among lesbian and bisexual women. Perspectives on sexual and reproductive health. 2005;37(1):6-12.
- 16. Lochner HJ, Maraqa NF. Sexually transmitted infections in pregnant women: integrating screening and treatment into prenatal care. Pediatric Drugs. 2018; 20:501-9.
- 17. Meites E, Gaydos CA, Hobbs MM, Kissinger P, Nyirjesy P, Schwebke JR, Secor WE, Sobel JD, Workowski KA. A review of evidence-based care of symptomatic trichomoniasis and asymptomatic Trichomonas vaginalis infections. Clinical infectious diseases. 2015;61(suppl\_8): S837-48.
- Kissinger PJ, Gaydos CA, Seña AC, Scott McClelland R, Soper D, Secor WE, Legendre D, Workowski KA, Muzny CA. Diagnosis and management of Trichomonas vaginalis: summary of evidence reviewed for the 2021 Centers for Disease Control and Prevention sexually transmitted infections treatment guidelines. Clinical Infectious Diseases. 2022;74(Supplement\_2): S152-61.
- 19. Kraus RF, Gruber MA. Neutrophils—From bone marrow to first-line defense of the innate immune system. Frontiers in immunology. 2021; 12:767175.
- Hiremath PS, Bannigidad P, Geeta S. Automated identification and classification of white blood cells (leukocytes) in digital microscopic images. IJCA special issue on "recent trends in image processing and pattern recognition" RTIPPR. 2010:59-63.
- 21. Nauseef WM. Human neutrophils≠ murine neutrophils: Does it matter? Immunological reviews. 2023314(1):442-56.
- 22. Teng TS, Ji AL, Ji XY, Li YZ. Neutrophils and immunity: from bactericidal action to being conquered. Journal of immunology research. 2017.
- 23. Fang FC. Antimicrobial reactive oxygen and nitrogen species: concepts and controversies. Nature Reviews Microbiology. 2004;2(10):820-32.
- 24. Zawrotniak M, Rapala-Kozik M. Neutrophil extracellular traps (NETs)-formation and implications. Acta Biochimica Polonica. 2013;60(3).
- 25. Nel JG, Theron AJ, Pool R, Durandt C, Tintinger GR, Anderson R. Neutrophil extracellular traps and their role in health and disease. South African Journal of Science. 2016;112(1-2):01-9.
- 26. Jones HR, Robb CT, Perretti M, Rossi AG. The role of neutrophils in inflammation resolution. InSeminars in immunology 2016; 28 (2):137-145). Academic Press.
- 27. Liew PX, Kubes P. The neutrophil's role during health and disease. Physiological reviews. 2019;99(2):1223-48.
- 28. Leliefeld PH, Wessels CM, Leenen LP, Koenderman L, Pillay J. The role of neutrophils in immune dysfunction during severe inflammation. Critical care. 2016; 20:1-9.
- 29. Rosales C. Neutrophils at the crossroads of innate and adaptive immunity. Journal of Leucocyte Biology. 2020;108(1):377-96.
- 30. Ferla M, Tasca T. The role of purinergic signaling in Trichomonas vaginalis infection. Current Topics in Medicinal Chemistry. 2021 Jan 1;21(3):181-92.
- 31. Bhakta SB, Moran JA, Mercer F. Neutrophil interactions with the sexually transmitted parasite Trichomonas vaginalis: implications for immunity and pathogenesis. Open biology. 2020;10(9):200192.
- 32. Mercer F, Ng SH, Brown TM, Boatman G, Johnson PJ. Neutrophils kill the parasite Trichomonas vaginalis using trogocytosis. PLoS biology. 2018;16(2): e2003885.
- 33. Ramírez-Ledesma MG, Romero-Contreras YJ, Rodríguez MC, Reyes-Cortes R, Cuéllar-Mata P, Avila EE. Trichomonas vaginalis triggers neutrophil extracellular traps reducing parasite integrity and growth. Parasitology Research. 2022;121(5):1355-67.
- 34. Bouchery T, Harris N. Neutrophil-macrophage cooperation and its impact on tissue repair. Immunology and cell biology. 2019;97(3):289-98.
- 35. de Andrade MF, de Almeida VD, de Souza LM, Paiva DC, Andrade CD, de Medeiros Fernandes TA. Involvement of neutrophils in Chagas disease pathology. Parasite immunology. 2018;40(12): e12593.
- 36. Kupani M, Pandey RK, Mehrotra S. Neutrophils and Visceral Leishmaniasis: Impact on innate immune response and cross-talks with macrophages and dendritic cells. Journal of Cellular Physiology. 2021;236(4):2255-67.
- 37. Zambrano F, Melo A, Rivera-Concha R, Schulz M, Uribe P, Fonseca-Salamanca F, Ossa X, Taubert A, Hermosilla C, Sánchez R. High presence of NETotic cells and neutrophil extracellular traps in vaginal discharges of women with vaginitis: an exploratory study. Cells. 2022;11(20):3185.

- Menezes CB, Tasca T. Trichomoniasis immunity and the involvement of the purinergic signaling. Biomedical journal. 2016;39(4):234-43.
- Riestra AM, de Miguel N, Dessi D, Simoes-Barbosa A, Mercer FK. Trichomonas vaginalis: lifestyle, cellular biology, and molecular mechanisms of pathogenesis. InLifecycles of Pathogenic Protists in Humans 2022; 541-617. Cham: Springer International Publishing.
- 40. Song HO, Lim YS, Moon SJ, Ahn MH, Ryu JS. Delayed human neutrophil apoptosis by Trichomonas vaginalis lysate. The Korean Journal of Parasitology. 2010;48(1):1.
- Frasson AP, Menezes CB, Goelzer GK, Gnoatto SC, Garcia SC, Tasca T. Adenosine reduces reactive oxygen species and interleukin-8 production by Trichomonas vaginalis-stimulated neutrophils. Purinergic signalling. 2017; 13:569-77.
- 42. Galego GB, Tasca T. Infinity war: Trichomonas vaginalis and interactions with host immune response. Microbial Cell. 2023;10(5):103.
- 43. Kusdian G, Gould SB. The biology of Trichomonas vaginalis in the light of urogenital tract infection. Molecular and biochemical parasitology. 2014;198(2):92-9.
- 44. Nemati M, Malla N, Yadav M, Khorramdelazad H, Jafarzadeh A. Humoral and T cell-mediated immune response against trichomoniasis. Parasite immunology. 2018;40(3): e12510.
- Alhussien MN, Dang AK. Potential roles of neutrophils in maintaining the health and productivity of dairy cows during various physiological and physiopathological conditions: a review. Immunologic Research. 2019; 67:21-38.
- 46. Obeagu EI, Obeagu GU, Ogunnaya FU. Deep vein thrombosis in pregnancy: A review of prevalence and risk factors. Int. J. Curr. Res. Chem. Pharm. Sci. 2023;10(8):14-21.
- 47. Obeagu EI, Obeagu GU. Molar Pregnancy: Update of prevalence and risk factors. Int. J. Curr. Res. Med. Sci. 2023;9(7):25-8.
- 48. Obeagu EI, Obeagu GU, Adepoju OJ. Evaluation of haematological parameters of pregnant women based on age groups in Olorunsogo road area of Ido, Ondo state. J. Bio. Innov11 (3). 2022:936-41.
- Obeagu EI, Adepoju OJ, Okafor CJ, Obeagu GU, Ibekwe AM, Okpala PU, Agu CC. Assessment of Haematological Changes in Pregnant Women of Ido, Ondo State, Nigeria. J Res Med Dent Sci. 2021;9(4):145-8.
- 50. Obeagu EI, Ezimah AC, Obeagu GU. Erythropoietin in the anaemias of pregnancy: a review. Int J Curr Res Chem Pharm Sci. 2016;3(3):10-8.
- 51. Sadik CD, Kim ND, Luster AD. Neutrophils cascading their way to inflammation. Trends in immunology. 2011;32(10):452-60.
- Hahn S, Hasler P, Vokalova L, van Breda SV, Lapaire O, Than NG, Hoesli I, Rossi SW. The role of neutrophil activation in determining the outcome of pregnancy and modulation by hormones and/or cytokines. Clinical & Experimental Immunology. 2019;198(1):24-36.
- 53. Bert S, Nadkarni S, Perretti M. Neutrophil-T cell crosstalk and the control of the host inflammatory response. Immunological Reviews. 2023;314(1):36-49.
- 54. Sabic D, Koenig JM. A perfect storm: fetal inflammation and the developing immune system. Pediatric research. 2020;87(2):319-26.
- 55. Edwards T, Burke P, Smalley H, Hobbs G. Trichomonas vaginalis: Clinical relevance, pathogenicity and diagnosis. Critical reviews in microbiology. 2016;42(3):406-17.
- 56. Lochner HJ, Maraqa NF. Sexually transmitted infections in pregnant women: integrating screening and treatment into prenatal care. Pediatric Drugs. 2018; 20:501-9.
- 57. Hou K, Wu ZX, Chen XY, Wang JQ, Zhang D, Xiao C, Zhu D, Koya JB, Wei L, Li J, Chen ZS. Microbiota in health and diseases. Signal transduction and targeted therapy. 2022;7(1):135.
- 58. Wu Z, Wang L, Li J, Wang L, Wu Z, Sun X. Extracellular vesicle-mediated communication within host-parasite interactions. Frontiers in immunology. 2019; 9:3066.
- 59. Yadav S, Verma V, Singh Dhanda R, Yadav M. Insights into the toll-like receptors in sexually transmitted infections. Scandinavian Journal of Immunology. 2021;93(1): e12954.
- Obeagu EI, Obeagu GU, Igwe MC, Alum EU, Ugwu OP. Neutrophil-Derived Inflammation and Pregnancy Outcomes.Newport International Journal Of Scientific And Experimental Sciences 4(2):10-19.https://doi.org/10.59298/NIJSES/2023/10.2.1111
- 61. Obeagu EI, Agreen FC. Anaemia among pregnant women: A review of African pregnant teenagers. J Pub Health Nutri. 2023; 6 (1). 2023;138.links/63da799664fc860638054562/Anaemia-among-pregnant-women-A-review-of-African-pregnant-teenagers.pdf.

47-54

- 62. Obeagu EI, Ezimah AC, Obeagu GU. Erythropoietin in the anaemias of pregnancy: a review. Int J Curr Res Chem Pharm Sci. 2016;3(3):10-8.links/5710fae108ae846f4ef05afb/ERYTHROPOIETIN-IN-THE-ANAEMIAS-OF-PREGNANCY-A-REVIEW.pdf.
- Obeagu EI, Adepoju OJ, Okafor CJ, Obeagu GU, Ibekwe AM, Okpala PU, Agu CC. Assessment of Haematological Changes in Pregnant Women of Ido, Ondo State, Nigeria. J Res Med Dent Sci. 2021 Apr;9(4):145-8.links/608a6728a6fdccaebdf52d94/Assessment-of-Haematological-Changes-in-Pregnant-Women-of-Ido-Ondo.pdf.
- 64. Obeagu EI, Obeagu GU. Sickle Cell Anaemia in Pregnancy: A Review. International Research in Medical and Health Sciences. 2023 Jun 10;6(2):10-3.http://irmhs.com/index.php/irmhs/article/view/111.
- 65. Jakheng SP, Obeagu EI. Seroprevalence of human immunodeficiency virus based on demographic and risk factors among pregnant women attending clinics in Zaria Metropolis, Nigeria. J Pub Health Nutri. 2022; 5 (8). 2022;137.links/6317a6b1acd814437f0ad268/Seroprevalence-of-human-immunodeficiency-virus-based-on-demographic-and-risk-factors-among-pregnant-women-attending-clinics-in-Zaria-Metropolis-Nigeria.pdf.
- 66. Obeagu EI, Obeagu GU, Chukwueze CM, Ikpenwa JN, Ramos GF. Evaluation of Protein C, Protein S and Fibrinogen of Pregnant Women with Malaria in Owerri Metropolis. Madonna University journal of Medicine and Health Sciences ISSN: 2814-3035. 2022 Apr 19;2(2):1-9.
- 67. Obeagu EI, Ikpenwa JN, Chukwueze CM, Obeagu GU. Evaluation of protein C, protein S and fibrinogen of pregnant women in Owerri Metropolis. Madonna University Journal of Medicine and Health Sciences ISSN: 2814-3035.
  2022 Apr 18;2(1):292-8.https://madonnauniversity.edu.ng/journals/index.php/medicine/article/view/57.
- 68. Obeagu EI, Obeagu GU, Adepoju OJ. Evaluation of haematological parameters of pregnant women based on age groups in Olorunsogo road area of Ido, Ondo state. J. Bio. Innov11 (3). 2022:936-41.
- 69. Obeagu EI. An update on utilization of antenatal care among pregnant Women in Nigeria. Int. J. Curr. Res. Chem. Pharm. Sci. 2022;9(9):21-6.DOI: 10.22192/ijcrcps.2022.09.09.003
- Okoroiwu IL, Obeagu EI, Obeagu GU. Determination of clot retraction in preganant women attending antenatal clinic in federal medical centre Owerri, Nigeria. Madonna University Journal of Medicine and Health Sciences ISSN: 2814-3035. 2022 Jul 22;2(2):91-7.
- 71. Obeagu EI, Hassan AO, Adepoju OJ, Obeagu GU, Okafor CJ. Evaluation of Changes in Haematological Parameters of Pregnant Women Based on Gestational Age at Olorunsogo Road Area of Ido, Ondo State. Nigeria. Journal of Research in Medical and Dental Science. 2021;9(12):462-.links/61b1e32f0c4bfb675178bfa7/Evaluation-of-Changes-in-Haematological-Parameters-of-Pregnant-Women-Based-on-Gestational-Age-at-Olorunsogo-Road-Area-of-Ido-Ondo-State-Nigeria.pdf.
- 72. Anyiam AF, Obeagu EI, Obi E, Omosigho PO, Irondi EA, Arinze-Anyiam OC, Asiyah MK. ABO blood groups and gestational diabetes among pregnant women attending University of Ilorin Teaching Hospital, Kwara State, Nigeria. International Journal of Research and Reports in Hematology. 2022 Jun 21;5(2):113-21.
- 73. Obeagu EI. Gestational Thrombocytopaenia. J Gynecol Women's Health. 2023;25(3):556163.links/64b01aa88de7ed28ba95fccb/Gestational-Thrombocytopaenia.pdf.
- 74. Jakheng SP, Obeagu EI, Abdullahi IO, Jakheng EW, Chukwueze CM, Eze GC, Essien UC, Madekwe CC, Madekwe CC, Vidya S, Kumar S. Distribution Rate of Chlamydial Infection According to Demographic Factors among Pregnant Women Attending Clinics in Zaria Metropolis, Kaduna State, Nigeria. South Asian Journal of Research in Microbiology. 2022;13(2):26-31.
- 75. Obeagu EI, Ogbonna US, Nwachukwu AC, Ochiabuto O, Enweani IB, Ezeoru VC. Prevalence of Malaria with Anaemia and HIV status in women of reproductive age in Onitsha, Nigeria. Journal of Pharmaceutical Research International. 2021 Feb 23;33(4):10-9.
- 76. Obeagu EI, Abdirahman BF, Bunu UO, Obeagu GU. Obsterics characteristics that effect the newborn outcomes. Int. J. Adv. Res. Biol. Sci. 2023;10(3):134-43.DOI: 10.22192/ijarbs.2023.10.03.016
- 77. Obeagu EI, Ogunnaya FU. PREGNANCYINDUCED HAEMATOLOGICAL CHANGES: A KEY TO MARTERNAL AND CHILD HEALTH. European Journal of Biomedical. 2023;10(8):42-3.links/64c890bddb38b20d6dad2c5c/PREGNANCY-INDUCED-HAEMATOLOGICAL-CHANGES-A-KEY-TO-MARTERNAL-AND-CHILD-HEALTH.pdf.
- 78. Ezeoru VC, Enweani IB, Ochiabuto O, Nwachukwu AC, Ogbonna US, Obeagu EI. Prevalence of Malaria with Anaemia and HIV status in women of reproductive age in Onitsha, Nigeria. Journal of Pharmaceutical Research International. 2021;33(4):10-9.
- 79. Okamgba OC, Nwosu DC, Nwobodo EI, Agu GC, Ozims SJ, Obeagu EI, Ibanga IE, Obioma-Elemba IE, Ihekaire DE, Obasi CC, Amah HC. Iron Status of Pregnant and Post-Partum Women with Malaria Parasitaemia in Aba Abia State, Nigeria. Annals of Clinical and Laboratory Research.

2017;5(4):206.links/5ea97df145851592d6a8acf2/Iron-Status-of-Pregnant-and-Post-Partum-Women-with-Malaria-Parasitaemia-in-Aba-Abia-State-Nigeria.pdf.

- 80. Eze RI, Obeagu EI, Edet FN. Frequency of Rh Antigen C And c among pregnant women in Sub-Urban area in Eastern Nigeria. Madonna Uni J Med Health Sci. 2021;1(1):19-30.
- Obeagu EI, Ofodile AC, Okwuanaso CB. A review of urinary tract infections in pregnant women: Risks factors. J Pub Health Nutri. 2023; 6 (1). 2023;137:26-35.links/63c3a9116fe15d6a571e8bba/A-review-of-urinary-tractinfections-in-pregnant-women-Risks-factors.pdf.
- 82. Obeagu EI, Obeagu GU, Musiimenta E. Post partum haemorrhage among pregnant women: Update on risks factors. Int. J. Curr. Res. Med. Sci. 2023;9(2):14-7.DOI: 10.22192/ijcrms.2023.09.02.003
- 83. Obeagu EI, Obeagu GU, Ogunnaya FU. Deep vein thrombosis in pregnancy: A review of prevalence and risk factors. Int. J. Curr. Res. Chem. Pharm. Sci. 2023;10(8):14-21.DOI: 10.22192/ijcrcps.2023.10.08.002
- 84. Jakheng SP, Obeagu EI, Jakheng EW, Uwakwe OS, Eze GC, Obeagu GU, Vidya S, Kumar S. Occurrence of Chlamydial Infection Based on Clinical Symptoms and Clinical History among Pregnant Women Attending Clinics in Zaria Metropolis, Kaduna State, Nigeria. International Journal of Research and Reports in Gynaecology. 2022 Aug 11;5(3):98-105.
- 85. Okorie HM, Obeagu EI, Eze EN, Jeremiah ZA. Assessment of some haematological parameters in malaria infected pregnant women in Imo state Nigeria. Int. J. Curr. Res. Biol. Med. 2018;3(9):1-4.DOI: 10.22192/ijcrbm.2018.03.09.001
- Onyenweaku FC, Amah HC, Obeagu EI, Nwandikor UU, Onwuasoanya UF. Prevalence of asymptomatic bacteriuria and its antibiotic susceptibility pattern in pregnant women attending private ante natal clinics in Umuahia Metropolitan. Int J Curr Res Biol Med. 2017;2(2):13-23.DOI: 10.22192/ijcrbm.2017.02.02.003
- Okoroiwu IL, Chinedu-Madu JU, Obeagu EI, Vincent CC, Ochiabuto OM, Ibekwe AM, Amaechi CO, Agu CC, Anoh NV, Amadi NM. Evaluation of Iron Status, Haemoglobin and Protein Levels of Pregnant Women in Owerri Metropolis. Journal of Pharmaceutical Research International. 2021 Apr 29;33(27A):36-43.
- 88. Obeagu EI, Njar VE, Obeagu GU. Infertility: Prevalence and Consequences. Int. J. Curr. Res. Chem. Pharm. Sci. 2023;10(7):43-50.
- 89. Emeka-Obi OR, Ibeh NC, Obeagu EI, Okorie HM. Evaluation of levels of some inflammatory cytokines in preeclamptic women in owerri. Journal of Pharmaceutical Research International. 2021 Aug 25;33(42A):53-65.
- 90. Obeagu EI, Faduma MH, Uzoma G. Ectopic Pregnancy: A Review. Int. J. Curr. Res. Chem. Pharm. Sci. 2023;10(4):40-4.DOI: 10.22192/ijcrcps.2023.10.04.004
- 91. Obeagu EI, Gamade SM, Obeagu GU. The roles of Neutrophils in pregnancy. Int. J. Curr. Res. Med. Sci. 2023;9(5):31-5.DOI: 10.22192/ijcrms.2023.09.05.005
- 92. Eze R, Obeagu EI, Nwakulite A, Okoroiwu IL, Vincent CC, Okafor CJ, Chukwurah EF, Chijioke UO, Amaechi CO. Evaluation of Copper Status and Some Red Cell Parameters of Pregnant Women in Enugu State, South Eastern Nigeria. Journal of Pharmaceutical Research International. 2021 May 29;33(30A):67-71.
- 93. Obeagu EI, Obeagu GU. Molar Pregnancy: Update of prevalence and risk factors. Int. J. Curr. Res. Med. Sci. 2023;9(7):25-8.DOI: 10.22192/ijcrms.2023.09.07.005
- 94. Obeagu EI, Bunu UO. Factors that influence unmet need for family planning. International Journal of Current Research in Biology and Medicine. 2023;8(1):23-7.
- 95. Ibebuike JE, Ojie CA, Nwokike GI, Obeagu EI, Nwosu DC, Nwanjo HU, Agu GC, Ezenwuba CO, Nwagu SA, Akujuobi AU. Barriers to utilization of maternal health services in southern senatorial district of Cross Rivers state, Nigeria. International Journal of Advanced Multidisciplinary Research. 2017;4(8):1-9.DOI: 10.22192/ijamr.2017.04.08.001
- 96. Emannuel G, Martin O, Peter OS, Obeagu EI, Daniel K. Factors Influencing Early Neonatal Adverse Outcomes among Women with HIV with Post Dated Pregnancies Delivering at Kampala International University Teaching Hospital, Uganda. Asian Journal of Pregnancy and Childbirth. 2023 Jul 29;6(1):203-11.http://research.sdpublishers.net/id/eprint/2819/.
- 97. Okorie HM, Obeagu EI, Eze EN, Jeremiah ZA. Assessment of coagulation parameters in malaria infected pregnant women in Imo state, Nigeria. International Journal of Current Research in Medical Sciences. 2018;4(9):41-9.DOI: 10.22192/ijcrms.2018.04.09.006
- Obeagu EI, Obeagu GU. Postpartum haemorrhage among women delivering through spontaneous vaginal delivery: Prevalence and risk factors. Int. J. Curr. Res. Chem. Pharm. Sci. 2023;10(8):22-6.DOI: 10.22192/ijcrcps.2023.10.08.003
- 99. Nwosu DC, Nwanjo HU, Obeagu EI, Ibebuike JE, Ezeama MC. Ihekireh. Changes in liver enzymes and lipid profile of pregnant women with malaria in Owerri, Nigeria. International Journal of Current Research and Academic Review. 2015;3(5):376-83.

47-54

100.Ibebuike JE, Ojie CA, Nwokike GI, Obeagu EI, Nwosu DC, Nwanjo HU, Agu GC, Ezenwuba CO, Nwagu SA, Akujuobi AU. Factors that influence women's utilization of primary health care services in Calabar Cros river state, Nigeria. Int. J. Curr. Res. Chem. Pharm. Sci. 2017;4(7):28-33.